

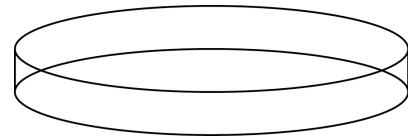
LAB ___: CLONING PAPER PLASMID

In this exercise you will use paper to simulate the cloning of a gene from one organism into a bacterial plasmid using a restriction enzyme digest. The plasmid (puc18 plasmid) can then be used to transform bacteria so that it now expresses a new gene and produces a new protein.

1. From the white paper, cut out the puc18 plasmid DNA in a long strip.

AAATCGTTTGC.....

2. Attach the ends together to make a loop to simulate the circular DNA of a plasmid.



3. From the green paper, cut out the Jellyfish *Glo* gene DNA in a long strip. Leave it as a straight strip. (This is a gene from a vertebrate not a bacterium, so it is not circular.)

GGATCGAAAGC.....

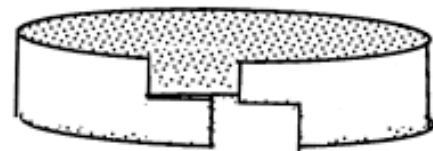
The start and stop sequences for transcribing the Jellyfish *GFP* or *Glo* gene are highlighted. These are needed to transcribe the gene properly when it is read.

In addition, the *Hind*III & *Eco*R1 restriction enzyme cutting sites (sequences of bases) are marked in **bold** on the Jellyfish *Glo* gene DNA. The two restriction enzymes and their respective restriction sites are listed below. These enzymes act as “molecular scissors” to cut the DNA at these sequences in the DNA:

Restriction enzyme	Recognition site (5'→3')
<i>Hind</i> III	A ↓ AGCT T T TCGA ↑ A
<i>Eco</i> RI	G ↓ AATT C C TTAA ↑ G

The six letter sequence represents the nitrogen base sequence that the enzyme recognizes, and ↑ represents the place where the DNA will be cut by the enzyme. For example, *Hind*III cuts between A and A whenever it encounters the six base sequence AAGCTT.

4. Cut the green Jellyfish DNA as if you have used the a restriction enzyme, *Hind*III. Be sure to leave “sticky ends.”
5. Also, cut the white puc18 plasmid DNA as if you have performed a restriction enzyme digest with *Hind*III. Be sure to leave “sticky ends.”



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6. Now you will incorporate the green Jellyfish *Glo* gene into the plasmid. Attach the sticky ends of the Jellyfish *Glo* gene to the sticky ends of the puc18 plasmid and seal with “molecular glue”, the enzyme ligase (scotch tape will be used in our lab).
7. You have successfully cloned a gene! You now have a single plasmid with a new gene and can use that to transform a single bacterium. The bacterium will now make green Jellyfish glow protein and will glow under black light.

QUESTIONS

1. What is a plasmid?

2. What are restriction enzymes used for in nature?

3. What is meant by a “sticky end”?

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4. Why did we cut both segments of DNA with the same restriction enzyme?

5. Why did we make sure to include the start and stop DNA sequences for the Jellyfish *Glo* gene in our cut segment?

6. What would have happened if we had cut both the Jellyfish *Glo* gene and puc18 plasmid with the EcoR1 restriction enzyme? Be sure to look on the paper DNA sequences to find the EcoR1 restriction enzyme cut sites.

7. If we want to now produce a lot of this Jellyfish *Glo* protein, what do we have to do after this first successful cloning to reach our goal?

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8. Scientists have successfully produced green fluorescent mice using this Jellyfish *GFP* gene. What do we now have to do to successfully use our cloned gene to transform mice. Go to the Web site <<http://www.rpc.msoe.edu/cbm2/gfp1.htm>> to see a photo of these transformed mice.

9. Scientists have successfully transformed bacteria with human genes. Describe one current use of the technology in medicine.

PRINT ON WHITE PAPER

Plasmid (puc18) DNA sequence

5'

3'

GAATCCGAAGCTCGGTACCCGGGGATCCTCTAGAGTCGACCTGCAGGCATGCAAGCTTGGCTACCGTGTACCTG
CTTAGGCTTCGAGCCATGGGCCCTAGGAGATCTCAGCTGGACGTCCGTACGTTCGAAACCGATGGCACATGGAC

Plasmid (puc18) DNA sequence

5'

3'

GAATCCGAAGCTCGGTACCCGGGGATCCTCTAGAGTCGACCTGCAGGCATGCAAGCTTGGCTACCGTGTACCTG
CTTAGGCTTCGAGCCATGGGCCCTAGGAGATCTCAGCTGGACGTCCGTACGTTCGAAACCGATGGCACATGGAC

Plasmid (puc18) DNA sequence

5'

3'

GAATCCGAAGCTCGGTACCCGGGGATCCTCTAGAGTCGACCTGCAGGCATGCAAGCTTGGCTACCGTGTACCTG
CTTAGGCTTCGAGCCATGGGCCCTAGGAGATCTCAGCTGGACGTCCGTACGTTCGAAACCGATGGCACATGGAC

PRINT ON GREEN PAPER

Chromosomal DNA (GFP gene) from Jellyfish: HindIII & EcoR1 restriction sites are marked in bold

5'

3'

GTGCGCG**AAGCTT**CCT**TACT**TCCAGAGC**GAATTC**TCTGGTCATTTTCTAGGCTATAT**ACT**TCTAA**AAGCTT**TTTCTG
CACGCGC**TTCGA**AGGAATGAGGTCTCG**CTTAAG**AGACCAGTAAAAGATCCGATATATGAAGAT**TTCGAAA**AGAC

GFP gene

Chromosomal DNA (GFP gene) from Jellyfish: HindIII & EcoR1 restriction sites are marked in bold

5'

3'

GTGCGCG**AAGCTT**CCT**TACT**TCCAGAGC**GAATTC**TCTGGTCATTTTCTAGGCTATAT**ACT**TCTAA**AAGCTT**TTTCTG
CACGCGC**TTCGA**AGGAATGAGGTCTCG**CTTAAG**AGACCAGTAAAAGATCCGATATATGAAGAT**TTCGAAA**AGAC

GFP gene

Chromosomal DNA (GFP gene) from Jellyfish: HindIII & EcoR1 restriction sites are marked in bold

5'

3'

GTGCGCG**AAGCTT**CCT**TACT**TCCAGAGC**GAATTC**TCTGGTCATTTTCTAGGCTATAT**ACT**TCTAA**AAGCTT**TTTCTG
CACGCGC**TTCGA**AGGAATGAGGTCTCG**CTTAAG**AGACCAGTAAAAGATCCGATATATGAAGAT**TTCGAAA**AGAC

GFP gene